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# (54) NOVEL GUANOSINE TRIPHOSPHATE (GTP)-BINDING PROTEIN CONJUGATE TYPE RECEPTOR PROTEINS

(57) The present invention provides a full-length cDNA encoding a rat G protein-coupled receptor protein, which was isolated by screening cDNA libraries originating from rat thalamus and hypothalamus. It also provides human cDNA corresponding to the rat cDNA. Use of these G protein-coupled receptor proteins makes it possible to screen ligands and also drug-candidate compounds capable of regulating signal transduction from receptors.

#### Description

### Technical Field

[0001] The present invention relates to a novel guanosine triphosphate binding protein-coupled receptor protein, a DNA encoding said protein, and a method for screening drug-candidate compounds using them.

#### Background Art

[0002] Many hormones and neurotransmitters regulate physiological functions through specific receptor proteins located on the cell membrane. Many of these receptor-proteins transduce signals into the cell by activating a guanosine triphosphate binding protein (occasionally referred to as "G protein" below) that is coupled to them. These receptor proteins are thereby named as G protein-coupled receptors. Since they have a common structure composed of seven transmembrane regions, they are also generally called "seven-transmembrane receptor proteins."

[0003] G protein-coupled receptors, which are expressed on the surface of cells *in vivo* and functioning cells of tissues, play an extremely important role as a target of molecules such as hormones, neurotransmitters, and biologically active compounds, which regulate the functions of these cells and tissues. Therefore, G protein-coupled receptor proteins have received great attention as targets in drug-development.

G protein-coupled receptors reported so far include, muscarinic acetylcholine receptors M1, M2, M3, and M4 (Peralta E.G. et al., EMBO J. 6:3923-3929 (1987)), muscarinic acetylcholine receptor M5 (Bonner T. I. et al., Neuron 1:403-410 (1988)), adenosine receptor A1 (Libert F. et al., Science 244:569-572 (1989)), α1A adrenoreceptor (Bruno J.F. et al., Biochem. Biophys. Res. Commun. 179:1485-1490 (1991)), β1 adrenoreceptor (Frielle T. et al., Proc. Natl. Acad. Sci. USA 84:7920-7924 (1987)), angiotensin receptor AT, (Takayanagi R. et al., Biochem. Biophys. Res. Commun. 183:910-916 (1992)), endothelin receptor ET<sub>A</sub> (Adachi M. et al., Biochem. Biophys. Res. Commun. 180:1265-1272 (1991)), gonadotropin releasing factor receptor (Kaker S.S. et al., Biochem. Biophys. Res. Commun. 189:289-295 (1992)), histamine receptor H<sub>2</sub> (Ruat M. et al., Proc. Natl. Acad. Sci. USA 87:1658-1672 (1992)), neuropeptide Y receptor Y1 (Larhammar D. et al., J. Biol. Chem. 267:10935-10938 (1992)), interleukin-8 receptor IL8RA (Holmes W.E. et al., Science 2563:1278-1280(1991)), dopamine receptor D<sub>1</sub> (Mahan L.C. et al., Proc. Natl. Acad. Sci. USA 87:2196-2200 (1990)), metabolic glutamate receptor mGluR1 (Masu M. et al., Nature 349:760-765 (1991)), and somatostatin receptor 30 SS<sub>1</sub> (Yamada Y. et al., Proc. Natl. Acad. Sci. USA 89:251-255) (for reference, Watson S. and Arkinstall S., The Gprotein Linked Receptor FactsBook, Academic Press (1994)). Examples of developed medicines aimed at G protein-coupled receptors are: terazosine hydrochloride (antihypertensive agent, α1 adrenoreceptor antagonist), atenolol (antiarrhythmia, β1 adrenoreceptor antagonist), dicyclomine hydrochloride (antispasmodic agent, acetylcholine receptor antagonist), ranitidine hydrochloride (drug for peptic ulcers, histamine receptor H2 antagonist), trazodone hydrochloride (antidepressant, serotonin receptor 5-HT1B antagonist), and buprenorphine hydrochloride (analgesic agent, opioid receptor k agonist) (for reference, Stadel J.M. et al., Trends Pharm. Sci. 18:430-437 (1997); Medicine Handbook 5th edition, Yakugyo-Jiho).

[0005] The hypothalamus, a part of the brain which governs a number of programs that trigger a particular response, contributes to the homeostasis of the internal environment by means of a variety of outputs, as the center of the autonomic nervous system. For instance, it releases hormones such as thyrotropic hormone releasing hormone, gonadotropic hormone releasing hormone, and growth hormone releasing hormone, and thereby regulates the entire endocrine system through the actions of these hormones on the specific receptors expressed in target cells. These outputs in the hypothalamus are thought to be mediated by receptors expressed in the hypothalamus and compounds reacting with them. Therefore, elucidation of the relationship between the compounds regulating the hypothalamus outputs and their specific receptors expressed in the hypothalamus is extremely important in developing novel medicines for the treatment of diseases arising from endocrine disorders.

#### Disclosure of the Invention

[0006] The present invention provides a novel G protein-coupled receptor protein expressed in the brain (in particular, thalamus and hypothalamus, etc.). It also provides a method for screening ligands and drug-candidate compounds using said receptor protein.

[0007] The inventors first selected a region highly conserved in known G protein-coupled receptor proteins, then designed primers corresponding to the region, and performed reverse transcriptase-polymerase chain reaction (RT-PCR) using mRNA obtained from rat thalamus and hypothalamus. Next, amplified clones were randomly selected, and their partial nucleotide sequences were determined. To remove known clones from the nucleotide sequence determined-clones, colony-hybridization was performed using as a probe, cDNA clones judged to be encoding a known G protein-coupled receptor protein by homology search. Negative clones that failed to hybridize with any probe were

selected. Using probes prepared based on the nucleotide sequence of the negative clones, the inventors screened cDNA libraries from rat thalamus and hypothalamus, and succeeded in isolating a full-length cDNA encoding a rat G protein-coupled receptor. They also succeeded in isolating a full-length human cDNA corresponding to the rat cDNA. Furthermore, northern blot analysis of the tissue specificity of the gene expression showed that these genes are specifically expressed in the brain.

[0008] These G protein-coupled receptors would be extremely useful in screening for ligands and compounds that regulate the signal transduction from receptors, which are anticipated to be utilized as novel medicines.

[0009] Thus, the present invention relates to novel G protein-coupled receptor proteins that are expressed in the brain, DNAs encoding them, and a method for screening ligands and compounds that are drug-candidate compounds using the proteins.

[0010] Specifically, the invention relates to:

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- (1) a guanosine triphosphate binding protein-coupled receptor protein comprising the amino acid sequence of SEQ
- ID NO: 1, or said amino acid sequence in which one or more amino acids are replaced, deleted, or added;
- (2) a guanosine triphosphate binding protein-coupled receptor protein comprising the amino acid sequence of SEQ
- ID NO: 20, or said amino acid sequence in which one or more amino acids are replaced, deleted, or added;
- (3) a guanosine triphosphate binding protein-coupled receptor protein encoded by a DNA which hybridizes with a DNA comprising the nucleotide sequence of SEQ ID NO: 2;
- (4) a guanosine triphosphate binding protein-coupled receptor protein encoded by a DNA which hybridizes with a DNA comprising the nucleotide sequence of SEQ ID NO: 21;
- (5) a partial peptide of the receptor protein as described in any one of (1) to (4);
- (6) a DNA encoding the receptor protein as described in any one of (1) to (4) or the partial peptide as described in (5);

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- (7) the DNA described in (6), wherein said DNA comprising the nucleotide sequence of SEQ ID NO: 2 or NO: 21;
- (8) a vector containing the DNA as described in any one of (5) to (7);
- (9) a transformant carrying the vector as described in (8);
- (10) a method of producing the receptor protein as described in any one of (1) to (4) or the partial peptide as described in (5), the method comprising culturing the transformant as described in (9);
- (11) a method of screening for a ligand of the receptor protein as described in any one of (1) to (4), the method comprising exposing a test compound to the receptor protein as described in any one of (1) to (4) or the partial peptide as described in (5), and selecting a compound that binds to said protein or partial peptide;
- (12) a method of screening for a compound that inhibits the binding between the receptor protein as described in any one of (1) to (4) and its ligand, the method comprising,
  - (a) exposing a ligand to the receptor protein as described in any one of (1) to (4) or the partial peptide as described in (5) in the presence of a test compound, and detecting the binding activity between said protein or partial peptide and the ligand, and.
  - (b) comparing the binding activity detected in (a) with that in the absence of the test compound, and selecting a compound that reduces the binding activity between said protein or partial peptide and the ligand;
- (13) a kit for screening a compound that inhibits the binding between the receptor protein as described in any one of (1) to (4) and its ligand, the kit comprising the receptor protein as described in any one of (1) to (4) or the partial peptide as described in (5); and
- (14) an antibody which binds to the receptor protein as described in any one of (1) to (4).

[0011] "G protein-coupled receptor protein" herein refers to a receptor protein that transduces intracellular signals by activating G proteins. "Ligand" refers to a natural compound capable of binding to a G protein-coupled receptor and inducing signal transduction. "Agonist" refers to a compound having a bioactivity similar to that of the ligands of G protein-coupled receptors, including both natural and artificially synthesized compounds. "Antagonist" refers to a compound capable of inhibiting the bioactivity of a ligand of a G protein-coupled receptor, including both natural and artificially synthesized compounds. "Protein" and "peptide" as used herein include their salts as well.

[0012] The present invention relates to a novel G protein-coupled receptor protein. The nucleotide sequence of the cDNA encoding rat G protein-coupled receptor "BG2" isolated in the present invention is shown as SEQ ID NO; 2, and the amino acid sequence of the "BG2" protein is shown as SEQ ID NO: 1. The nucleotide sequence of the cDNA encoding human G protein-coupled receptor "BG2" isolated herein is shown as SEQ ID NO: 21, and the amino acid sequence of the human "BG2" protein is shown as SEQ ID NO: 20.

[0013] Rat "BG2" protein has 26%, 25%, and 29% homology to known G protein-coupled receptors; bovine muscarinic acetylcholine receptor M3 protein (Lee P.H. et al., Biochim. Biophys. Acta 1223:151-154 (1994)), human mus-

carinic acetylcholine receptor M5 protein (Bonner T.I. et al., Neuron 1:403-410 (1988)), and mouse  $\alpha$ 2A adrenoreceptor (Link R. et al., Mol. Pharmacol. 42:16-27 (1992)), respectively. The result of the hydrophobicity plot analysis showed that rat "BG2" protein contains hydrophobic regions (seven transmembrane regions) characteristic to G protein-coupled receptors. In addition, the coding region of the rat "BG2" cDNA has a size of approximately 1.2 kb, a size similar to that of the known G protein-coupled receptors.

[0014] Human "BG2" protein has 32%, 28%, and 27% homology to known G protein-coupled receptors; human  $\alpha$ -2C-1 adrenoreceptor (Regan J.W. et al., Proc. Natl.Acad.Sci. USA 85:6301-6305 (1988)), mouse  $\beta$ -1 adrenoreceptor (Jasper J.R. et al., Biochim. Biophys. Acta 1178:307-309 (1993)), and human muscarinic acetylcholine receptor M3 protein (Peralta E.G. et al., EMBO J 6:3923-3929 (1987)), respectively.

[0015] These results suggest that the "BG2" proteins belong to the G protein-coupled receptor family, which further suggests that they participate in signal transduction through the activation of G proteins upon ligand binding.

**[0016]** Furthermore, the result of a northern blot analysis showed that the genes encoding the "BG2" proteins are specifically expressed in the brain. *In situ* hybridization detected a strong expression of the rat "BG2" gene in the hippocampus and spinal cord, and also in the hypothalamus, thalamus, and cerebellum.

[0017] The hippocampus plays an important role in memory and learning, the cerebellum regulates the body motions, and the hypothalamus serves as the center of the autonomic nervous system. Thus, the "BG2" proteins are assumed to be involved in the regulation of these functions. Therefore, the "BG2" proteins and genes, or an agonist or antagonist that can regulate the "BG2" protein function(s) can be used in the treatment of disabilities in memory and learning, or the control of the autonomous nervous system, such as regulation of, blood pressure, digestion, body temperature, and food-intake etc.

[0018] "BG2" protein may be prepared as natural protein, and also as recombinant protein by using recombinant DNA technology. A natural protein may be prepared, for instance, by extracting tissues such as the thalamus and hypothalamus, speculated to express "BG2" protein, and performing immunoaffinity chromatography using anti-"BG2" antibody as described later on. On the other hand, the recombinant protein can be prepared by culturing transformant cells carrying DNA encoding "BG2" protein as described later on.

[0019] One skilled in the art can prepare an altered protein having a function (transduction of intracellular signals through G protein activation) equivalent to that of the natural protein by introducing modifications such as replacement of any amino acid contained in the natural rat or human "BG2" protein (SEQ ID NO: 1, or SEQ ID NO: 20, respectively) according to known methods. Mutations of amino acids in a protein may occur naturally. The G protein-coupled receptor protein of the present invention includes such mutant proteins having an amino acid sequence altered by replacement, deletion or addition, having a function equivalent to that of the natural protein. The methods of altering amino acids, known to one skilled in the art, include, the Kunkel method (Kunkel T.A. et al., Methods Enzymol. 154:367-382 (1987)), double primer method (Zoller M.J. and Smith M., Methods Enzymol. 154:329-350 (1987)), cassette mutation (Wells et al., Gene 34:315-323 (1985)), and megaprimer method (Sarkar G. and Sommer S.S., Biotechniques 8:404-407 (1990)). The number of mutated amino acids in a functionally equivalent protein is generally not more than 10% of all the amino acids, favorably not more than 10 amino acids, and more favorably not more than 3 amino acids (for instance, one amino acid).

[0020] One skilled in the art can also use hybridization techniques (Hanahan D. and Meselson M., Methods Enzymol. 100:333-342 (1983); Benton W.D. and Davis R.W., Science 196:180-182 (1977)) to isolate a highly homologous gene from various other species based on the sequence of the rat or human "BG2" cDNA (SEQ ID NO: 2, or SEQ ID NO; 21, respectively) or a part of it, and use the isolated DNA to obtain a protein a functionally equivalent to these "BG2" proteins. Thus, it is possible for one skilled in the art to prepare a protein functionally equivalent to the rat or human "BG2" proteins, which is encoded by a DNA that hybridizes to the rat or human "BG2" cDNA. The G protein-coupled receptor protein of the present invention includes these proteins as well. Species used for isolation of a functionally equivalent protein include mouse, rabbit, sheep, bovine, dog, and pig, for instance, and particularly the brain tissues such as thalamus and hypothalamus are suitable.

[0021] DNA encoding a functionally equivalent protein to the rat or human "BG2" proteins has usually high homology to the nucleotide sequence of the rat or human "BG2" cDNA (SEQ ID NO: 2, and NO: 21). High homology generally means a sequence identity of not less than 70%, favorably not less than 80%, and more favorably not less than 90% at the nucleotide level. The sequence identity can be determined by the FASTA program.

[0022] Hybridization to isolate a DNA having high homology to the rat or human "BG2" cDNA is usually performed in "6x SSC, 40% formamide, at 25°C" followed by a wash in "1x SSC at 55°C," favorably in "6x SSC, 40% formamide, at 37°C" followed by a wash in "0.2x SSC at 55°C," and more favorably in "6x SSC, 50% formamide, at 37°C" followed by a wash in "0.1x SSC at 62°C." One skilled in the art may easily obtain a hybridization condition similar to the above conditions by appropriately selecting factors such as the dilution rate of SSC, the formamide concentration, and temperature.

[0023] The present invention also includes a partial peptide of the above-described G protein-coupled receptor protein. The partial peptide of the present invention includes, for instance, those corresponding to the N-terminal region of

the G protein-coupled receptor protein, which can be utilized to prepare an antibody. The partial peptide of the invention has a length of at least 15 amino acids, and favorably 20 amino acids.

Furthermore, the present invention relates to a DNA encoding the G protein-coupled receptor protein of the invention as described above or its partial peptide. The DNA encoding the G protein-coupled receptor protein of the invention or its partial peptide includes cDNA, genomic DNA, and synthetic DNA, but is not limited as far as it encodes the protein or the peptide. cDNA encoding the G protein-coupled receptor of the invention can be obtained by screening a cDNA library from a tissue expressing the receptor (for instance, thalamus and hypothalamus) by using as a <sup>32</sup>Plabeled probe, the cDNA as described in SEQ ID NO: 2 or NO: 21, or a part of it, complementary RNA to the DNA, or a synthetic oligonucleotide comprising a part of the cDNA. Alternatively, cDNA may be cloned by synthesizing an oligonucleotide corresponding to the nucleotide sequence of the cDNA, and amplifying cDNA from an appropriate tissue (such as thalamus and hypothalamus) by PCR. Genomic DNA can be obtained by screening a genomic library by hybridization using as a <sup>32</sup>P-labled probe, the cDNA as described in SEQ ID NO: 2 or NO: 21, or a part of it, complementary RNA to the DNA, or a synthetic oligonucleotide comprising a part of the cDNA. Alternatively, it may be cloned by synthesizing an oligonucleotide corresponding to the nucleotide sequence of the cDNA, and amplifying genome DNA by PCR. Synthetic DNA can be prepared by chemically synthesizing oligonucleotides comprising a part of the nucleotide sequence of SEQ ID NO: 2 or NO: 21, annealing them into a double strand, and ligating them using DNA ligase (Khorana H.G. et al., J. Biol. Chem. 251:565-570 (1976); Goeddel D.V. et al., Proc. Natl. Acad. Sci. USA 76:106-110 (1979)).

[0025] These DNA can be used for producing recombinant proteins. Namely, it is possible to prepare the G protein-coupled receptor protein of the invention as a recombinant protein by inserting a DNA encoding the receptor protein (DNA as described in SEQ ID NO: 2 or NO: 21, for instance) into an appropriate expression vector, culturing a transformant obtained by introducing the vector into an appropriate cell, and purifying the expressed protein. Since the G protein-coupled receptor protein of the invention is a receptor protein, it is possible to prepare it in a form expressed on the cell membrane.

[0026] Specifically, if the host is *Escherichia coli*, the plasmid vectors such as pET-3 (Rosenburg A.H. et al., Gene 56:125-135 (1987)) and pGEX-1 (Smith D.B. and Johnson K.S., Gene 67:31-40 (1988)) may be used. *E.coli* can be transformed by the Hanahan method (Hanahan D., J. Mol. Biol. 166:557-580 (1983)), electroporation (Dower W.J. et al., Nucleic Acids Res. 16:6127-6145 (1988)), and such. If the host is fission yeast (Schizosaccharomyces pombe), a plasmid vector such as pESP-1 (Lu Q. et al., Gene 200:135-144 (1997)) is used. Yeast can be transformed by spheroplast fusion (Beach D. and Nurse P., Nature 290:140 (1981)), and lithium acetate method (Okazaki K. et al., Nucleic Acids Res. 18:6485-6489 (1990)), etc.

[0027] If the host is a mammalian cell, such as Chinese Hamster ovary-derived CHO cells and human HeLa cells, vectors such as pMSG (Clontech) are used. Recombinant DNA is introduced into mammalian cells by calcium phosphate method (Graham F.L. and van derEb A.J., J. Virology 52:456-467 (1973)), DEAE-dextran method (Sussman D.J. and Milman G., Mol. Cell. Biol. 4:1641-1643 (1984)), lipofection (Felgner P.L. et al., Proc. Natl. Acad. Sci. USA 84:7413-7417 (1987)), and electroporation (Neumann E. et al., EMBO J. 1:841-845 (1982)), etc. If the host is an insect cell, a baculovirus vector such as pBacPAK8/9 (Clontech) can be used. Transformation of insect cells is done by the methods described in the literature (BioTechnology 6:47-55 (1980)).

[0028] Recombinant proteins expressed in host cells can be purified by known methods. The proteins can also be synthesized as fusion proteins tagged with histidine residues at the N-terminus, or fused to glutathione-S-transferase (GST), and purified by using their binding ability toward a metal chelating resin, or a GST affinity resin (Smith M.C. et al., J. Biol. Chem. 263:7211-7215 (1988)), respectively. For instance, when the vector pESP-1 is used, the protein of interest is synthesized as a fusion protein with GST, which can be purified using GST affinity resin. The fusion protein may be digested with thrombin, or blood coagulating factor Xa to liberate the protein of interest.

[0029] Moreover, DNA encoding the G protein-coupled receptor protein of the present invention can be used in gene therapy of diseases that arise from a mutation of the protein. When used in gene therapy, the DNA can be introduced into human cells using retrovirus vectors (Danos O. and Mulligan R.C., Proc. Natl. Acad. Sci. USA 85:6460-6464 (1988); Dranoff et al., Proc. Natl. Acad. Sci. USA 90:3539-3543 (1993)), or adenovirus vectors (Wickham T.J. et al., Cell 73:309-319 (1993)), etc. To administer the vector to patients, transplantation of bone marrow, subcutaneous injection, and intravenous injection can be used (Asano S., Protein Nucleic acid and Enzyme 40:2491-2495 (1995)).

[0030] Furthermore, the present invention relates to an antibody that is capable of binding to the G protein-coupled receptor protein of the invention. Antibodies against the G protein-coupled receptor protein can be prepared by known methods in the art (for instance, refer to Shin-Seikagaku-Jikken-Kouza I: Protein I 389-406, Tokyo-Kagaku-Doujin). For instance, polyclonal antibodies are prepared as follows. An appropriate dose of the above protein or its partial peptide is administered into immune animals such as rabbits, guinea pigs, mice, or chickens. Administration may be performed together with the adjuvant (such as FIA or FCA) that promotes antibody production, and usually performed every couple of weeks. The titer of antibodies can be increased by performing multiple immunizations. After the final immunization, antisera are obtained by withdrawing blood from immune animals. Polyclonal antibodies are purified from antisera by

ammonium sulfate precipitation, fractionation by anion exchange chromatography, or affinity chromatography with either Protein A or immobilized antigen. Monoclonal antibodies are prepared as follows. The G protein-coupled receptor proteins of the invention or its partial peptide is administered into immune animals as described above. After the final immunization, their spleens or lymph nodes are excised. Then, antigen-producing cells are recovered from the spleens or the lymph nodes, and fused with myeloma cells using polyethylene glycol etc. to produce hybridomas. Desired hybridomas are selected by screening, and their culture supernatant is used to prepare monoclonal antibodies. Monoclonal antibodies can be purified by ammonium sulfate precipitation, fractionation by anion exchange chromatography, or affinity chromatography with either Protein A or immobilized antigen. Antibodies prepared thereby can be used not only in affinity purification of the G protein-coupled receptor protein of the invention, but also for the diagnosis or antibody treatment of diseases arising from the abnormal expression of the receptors, or detection of the expression level of the receptors.

[0031] If used for antibody treatment, humanized antibodies or human antibodies are preferable. Humanized antibodies, in case of mouse-human chimeric antibodies, are prepared by isolating the gene encoding the antibody against the G protein-coupled receptor protein from the producing mouse cells, replacing the constant region of the H chain of the antibody with that of the human IgE, and introducing it into mouse myeloma J558L cells (Neuberger M.S. et al., Nature 314:268-270 (1985)). Human antibodies can be prepared by immunizing mice, whose immune system is replaced with that of human, with the G protein-coupled receptor protein.

Furthermore, the present invention relates to a method of screening ligands of the G protein-coupled receptor protein of the invention. The method includes such processes as exposing a test compound to the G protein-coupled receptor protein or its partial peptide, and selecting compounds that are capable of binding to the proteins or the peptide. Compounds to be tested include known compounds such as acetylcholine, adenosine, adrenaline, noradrenaline, angiotensin, bombesin, bradykinin, C5a., anaphylatoxin, calcitonin, cannabinoids, chemokines, cholecystokinin, dopamine, endothelin, formylmethionylpeptide, GABA, galanin, glucagon, glutamate, glycopeptide hormone, histamine, 5-hydroxytryptophan, leucotriene, melanocortin, neuropeptide Y, neurotensin, odorant, opioid peptide, opsin, parathyroid hormone, platelet activating factor, prostanoid, somatostatin, tachykinin, thrombin, thyrotropin releasing hormone, vasopressin, and oxytocin (Watson S. and Arkinstall S., G protein Linked Receptor FactsBook, Academic Press (1994)), and also other purified proteins, expressed products of genes (including libraries), extracts of tissues or cells in which the ligand is stipulated to be expressed (the brain, thalamus, and hypothalamus etc.), and the culture medium of the cells. The G protein-coupled receptor proteins may be used in a form expressed in desired cells (including transformants genetically engineered to express the proteins) or on the cell surface, in a form of the membrane fractions of the cells, or in a form bound to an affinity column. If necessary, test compounds may be labeled appropriately. Methods for labeling include radioisotope labeling, and fluorescence labeling, but not limited thereto. The binding between the G protein-coupled receptor proteins and test compounds can be examined by detecting the label added to the compound (for instance, measuring the radioactivity or fluorescence intensity), or using as an index, intracellular signaling triggered by the compound binding to the G protein-coupled receptor protein (such as G protein activation, the change in the concentration of Ca2+ or cAMP, phospholipase C activation, and the change in pH). Specific methods can be employed as described in the literatures (Cell Calcium 14:663-671 (1993); Analytical Biochemistry 226:349-354 (1995); J. Biol. Chem. 268:5957-5964 (1993); Cell 92:573-585 (1998); Nature 393:272-273 (1998)), and unexamined published Japanese patent application (JP-A) No. Hei 9-268). Alternatively, the binding may be detected by measuring the activity of the reporter gene using two-hybrid system (Zervos et al., Cell 72:223-232 (1994); Fritz et al., Nature 376:530-533 (1995)).

[0033] The present invention also relates to a method of screening for a compound which can inhibit the binding between the G protein-coupled receptor proteins of the invention and their ligands. The method includes processes of (a) exposing the ligand to the G protein-coupled receptors or their partial peptides in the presence of test compound, and detecting the binding ability between the ligand and the proteins or peptides, and (b) comparing the ability detected in (a) with that in the absence of the compound, and selecting compounds which are capable of decreasing the binding ability. Compounds to be tested include proteins, peptides, non-peptide compounds, artificially synthesized compounds, extracts of tissues and cells, sera, but not limited thereto. The G protein-coupled receptor proteins may be used in a form expressed in desired cells (including transformants genetically engineered to express the proteins) or on the cell surface, in a form of the membrane fractions of the cells, or in a form bound to an affinity column. If necessary, tested compounds may be labeled appropriately. Methods for labeling include radioisotope labeling, and fluorescence labeling, but not limited thereto. The binding between the G protein-coupled receptor proteins and test compounds can be examined by detecting the label added to the compound (for instance, measuring the radioactivity or fluorescence intensity), or using intracellular signaling, as an index, that are triggered by the compound binding to the G protein-coupled receptor protein (such as G protein activation, the change in the concentration of Ca<sup>2+</sup> or cAMP, phospholipase C activation, and the change in pH). Specific methods can be employed as described in the literatures (Cell Calcium 14:663-671 (1993); Analytical Biochemistry 226:349-354 (1995); J. Biol. Chem. 268:5957-5964 (1993); Cell 92:573-585 (1998); Nature 393:272-273 (1998)), and JR-A Hei 9-268). If the result of the detection showed that the binding

activity in the presence of a test compound is lower than that in the absence of the compound, the compound is judged to be capable of inhibiting the binding between the G protein-coupled receptors or their partial peptides and their ligands. These compounds include those capable of triggering the intracellular signaling through binding to the G protein-coupled receptor (agonist), and those not having such activity (antagonist). Agonists have similar bioactivities to those of the ligands of the G protein-coupled receptors. On the other hand, antagonists inhibit the bioactivities of the ligands. Therefore, these agonists and antagonists are useful as medicinal components for treatment of diseases arising from disorders in the signaling pathway mediated by the G protein-coupled receptors.

[0034] Furthermore, the present invention relates to a screening kit for compounds that inhibit the binding between the G protein-coupled receptor proteins and their ligands. The G protein-coupled receptor proteins or their partial peptides may be in a form expressed in desired cells (including transformants genetically engineered to express the protein) or on the cell surface, in a form of membrane fractions of the cell, or in a form bound to an affinity column. Components of the kit of the invention may include, other than the above described receptor protein samples, ligand samples (both labeled and unlabeled), and buffers for the reaction between the ligand and the receptor protein, and wash solutions. Labels to be added to the ligands include radioisotope and fluorescence, for instance. The kit of the invention can be used as described in JP-A Hei 9-268.

#### Brief Description of the Drawings

#### [0035]

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Figure 1 shows the hydrophobicity plot of the mouse BG2 protein. The seven hydrophobic regions (transmembrane regions) that are characteristics of the G protein-coupled receptor proteins are indicated by the numbers from 1 to 7. The numbers in the bottom indicate those of the amino acid residues in the "BG2" protein.

Figure 2 shows the result of northern blot analysis of the tissue specific expression of the human and mouse BG2 genes.

Figure 3 shows the result of *in situ* hybridization analysis of the location of the mouse "BG2" gene expression in the brain.

Figure 4 shows the result of *in situ* hybridization analysis of the location of the mouse "BG2" gene expression in the spinal cord. "Sense" and "Antisense" indicate the results using sense RNA probe (not hybridizing with mRNA; negative control), and antisense RNA probe (hybridizing with mRNA), respectively.

### Best Mode for Carrying out the Invention

[0036] The present invention is illustrated in detail below with reference to examples, but is not to be construed as being limited thereto.

Example 1: Isolation of a gene encoding a rat G protein-coupled receptor

[0037] The G protein-coupled receptors share a characteristic structure composed of seven transmembrane regions, and the amino acid sequences of the transmembrane regions and the adjacent regions are well conserved. The present inventors first compared the nucleotide sequences of the second and the seventh transmembrane domains, which are highly conserved, with known G protein-coupled receptors: mouse neuropeptide Y receptor Y1 (GenBank Accession Number Z18280), rat Y1 (Z11504), human Y1 (M84755), mouse neuropeptide Y receptor Y4 (U40189), rat Y4 (Z68180), human Y4 (Z66526), and mouse neuropeptide Y receptor Y6 (U58367), and synthesized novel sense and antisense primers, as described in SEQ ID NO: 3 and NO: 4, respectively.

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[0038] Next, single stranded cDNA was synthesized from poly(A)\*RNA prepared from rat thalamus and hypothalamus using the RNA-PCR kit (TaKaRa), and PCR was performed using the two primers. Specifically, poly(A)\*RNA was prepared from rat thalamus and hypothalamus using Fasttrack 2.0 kit (Invitrogen). Then, 75 ng of the poly(A)\*RNA was used to synthesize complementary DNA according to the protocol accompanying the RNA-PCR kit (TaKaRa). PCR amplification was performed using all the cDNA. The reaction mixture comprising each 0.15 mM dNTPs, 1.5 mM MgCl<sub>2</sub>, 0.025 U/μl rTaq polymerase (TaKaRa), each 0.5 μM degenerated primer Fg (SEQ ID NO: 3) and Rb (SEQ ID NO: 4), and enzyme accompanying 10x PCR buffer was prepared making a total of 130 μl, and aliquoted into six 20 μl fractions. PCR was performed with the Pertier thermal cycler PTC200 (MJ Research) under conditions as follows: a single cycle of 94°C for 2 min, followed by 35 cycles of 94°C for 30 sec, 48°C for 1 min, 72°C for 1 min 30 sec, and then a single cycle of 72°C for 8 min. After PCR, the six reaction-solutions were combined, and the amplified products were purified using the Wizard PCR purification kit (Promega), and then eluted with 30 μl TE. 2 μl of the TE eluate was used for cloning into the pCR2.1 vector of the TOPO TA cloning kit (Invitrogen). XL1-Blue cells were used as the host cell and transformed using the *E.coli* pulser (BioRad). From the resulting transformants, 5,760 colonies having white or light blue

color were randomly selected using the gene library construction system BioPick (BioRobotics), and inoculated into fifteen 384-well plates containing LB media supplemented with 100  $\mu$ g/ml of ampicillin. Clones were cultured at 37°C overnight, and replica plated onto a filter on top of a LB agar plate containing 100  $\mu$ g/ml ampicillin and 25% glycerol, and another filter on top of a LB agar plate containing 100  $\mu$ g/ml ampicillin, for preparing a glycerol stock, and colony hybridization, respectively, using a gene library replicating system BioGrid (BioRobotics).

Since the obtained PCR clones were expected to contain multiple overlapping clones of the NPY receptor cDNA, 80 clones out of the 5,760 clones were randomly selected, and their nucleotide sequences were partially determined. To determine the nucleotide sequence, plasmid DNA purified by the plasmid automatic isolating system PI100sigma (Kurabo) was used as a template. The sequence reactions were performed using the dye-primer-cycle sequencing kit FS (Perkin Elmer), and the reaction products were separated by electrophoresis using the DNA sequencer 377 (Perkin Elmer). The homology search of the obtained sequence using the blast program of the Wisconsin package (Genetic Computer Group) showed that 29 out of the 80 clones were the cDNAs encoding the coiled-coil like protein 1 (GeneBank Accession Number U79024) while 17 clones were those of the neuropeptide Y receptor Y1 (Z11504). Then, these two cDNA fragments were used as a probe for hybridization with the filters containing a library of the degenerated PCR amplified fragments. Probes were prepared by amplifying the insert of the respective clones by PCR, purifying the products using the Wizard PCR purification kit (Promega), and labeling them with  $[\alpha^{-32}P]$  dCTP using the Prime-It II random primer labeling kit (Stratagene). Colony hybridization was performed according to the standard method (Sambrook et al., Molecular Cloning: A laboratory manual 2<sup>nd</sup> edition (1989)). Colonies that were negative for either the coiled-coil like protein 1 or the neuropeptide Y receptor Y1 were selected and their partial nucleotide sequences were determined. For DNA sequencing, the insert fragment of each clone was amplified by PCR from the culture medium, purified using the PCR product purification kit (Amersham), and used as a template. The sequence reactions were performed using the dye-primer-cycle sequencing kit FS (Perkin Elm. 1), and the reaction products were separated by electrophoresis using the DNA sequencer 377 (Perkin Elmer). The obic ned sequences were analyzed by the homology search using the blast program of the Wisconsin package (Genetic Computer Group), and, as a result, a clone which has significant homology to the muscarinic acetylcholine receptor M5 (GeneBank Accession Number M22926) was found. The clone has been deposited at the National Institute of Bioscience and Human-Technology, Agency of Industrial Science and Technology.

Name of the depositary institution: National Institute of Bioscience and Human-Technology, Agency of Industrial Science and Technology, MITI.

Address of the depositary institution: 1-1-3 Higashi, Tsukuba, Ibaraki 305-8566, Japan.

Date of deposit: 12/25/1997.

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Accession Number: FERM BP-6575

[0040] Next, in order to isolate the full-length cDNA of the gene, cDNA libraries were prepared from rat thalamus and hypothalamus. cDNA was synthesized according to the protocol accompanying the cDNA synthesizing kit (Stratagene), and the vector pEF1x and the host XL1-blue MRF' (Stratagene) were used. The pEF1x is a refined derivative of the pcDNA3 (Invitrogen) prepared as follows.

(1) Preparation of the human EF1a promoter (GeneBank Accession Number J04617)

[0041] PCR was performed using human genomic DNA with primers (SEQ ID NO: 6/ CGAGGATCCGTGAGGCTC-CGGTGCCCGTC; SEQ ID NO: 7/ CGGGTAAGCTTCACGACACCTGAAATGGAAGA). The products were digested with BamHI (TaKaRa) and HindIII (TaKaRa), and subcloned into the plasmid vector pUC19 (TaKaRa). The resulting plasmid was digested with XhoI, blunt-ended with the Klenow fragment (TaKaRa), and self-ligated using the DNA ligation kit (TaKaRa). The resulting plasmid was digested with BamHI and HindIII, and the insert was recovered.

(2) Alteration of the pcDNA3

[0042] The pcDNA3 was digested with Mlul (TaKaRa), blunt-ended with the Klenow fragment (TaKaRa), and self-ligated using the DNA ligation kit. The resulting plasmid was digested with AfIIII (New England Biolabs) and Smal (TaKaRa), blunt-ended with the Klenow fragment (TaKaRa), and self-ligated using the DNA ligation kit. Then, the obtained plasmid was digested with BgIII (TaKaRa) and HindIII, and the fragment depleted of the CMV promoter was recovered, and ligated with the insert fragment as described in (1) using the DNA ligation kit to construct the pEF1x.

[0042] Next, the guelectide sequence of the cDNA fragment was used to synthesize plinging cleating probe (SEQ ID).

[0043] Next, the nucleotide sequence of the cDNA fragment was used to synthesize oligonucleotide probe (SEQ ID NO: 8/ CCTTCTGCATCCCATTGTACGTACC), and multiple clones were obtained from the above cDNA libraries from rat thalamus and hypothalamus according to the protocol of the gene trapper cDNA positive selection system (GIBCO BRL). Then, colony hybridization was performed using the cDNA insert of the above isolated clone (FERM BP-6574)

as a probe, and a positive clone was obtained. This clone has been deposited at the National Institute of Bioscience and Human-Technology, Agency of Industrial Science and Technology.

Name of the depositary institution: National Institute of Bioscience and Human-Technology, Agency of Industrial Science and Technology, MITI.

Address of the depositary institution: 1-1-3 Higashi, Tsukuba, Ibaraki 305-8566, Japan.

Date of deposit: 12/25/1997. Accession Number: FERM BP-6574

[0044] The insert fragment of the clone was of 2.7 kb. Plasmid DNA was prepared by the QIAprep Midi Kit (QIA-GEN), and the complete nucleotide sequence was determined using the shotgun cloning method (Sambrook et al.., Molecular Cloning: A laboratory manual 2<sup>nd</sup> edition (1989)). cDNA fragmentation was performed using the closed sonifier biomaterial treating system Biorupter (Tousou Denki), and the DNA fragments were separated by electrophoresis on a 2% agarose gel. Fragments of around 0.6 kb were purified using the gene clean spin kit (bio101), blunt-ended with T4 DNA polymerase (TaKaRa), and cloned into the Hincll-BAP treated pUC118 vector, XL1-Blue was used as a host cell, and transformed using the *E.coli* pulser (BioRad). The obtained shotgun clones were sequenced using the dye-primer cycle sequencing kit FS (Perkin Elmer), or the dye-terminator cycle sequencing kit (Perkin Elmer). The resulting sequences were combined and edited to get the complete nucleotide sequence using the DNA sequencing software Sequencher (Hitachi Software). The complete nucleotide sequence is composed of 2700 bp, and turned out to be encoding a protein of 413 amino acids (SEQ ID NO: 5). Because there is a stop codon in the 5' region of the open reading frame, the cDNA is supposed to include the entire coding region (SEQ ID NO: 2). When this sequence was translated into the amino acid sequence, the hydrophobicity plot identified seven transmembrane regions from 1 to 7 (Figure 1).

[0045] In addition, the open reading frame size was approximately 1.2 kb, which is similar to that of the known G protein-coupled receptors. G protein-coupled receptor proteins have common features in their amino acid sequences, and thus form a protein family. As a result of the homology search using the amino acid sequence encoded by the isolated cDNA, the encoded protein was found to be a novel receptor protein having a homology of 26%, 25%, and 29% to known G protein-coupled receptors: bovine muscarinic acetylcholine receptor M3 protein (Lee P.H. et al., Biochim. Biophys. Acta 1223:151-154 (1994)), human muscarinic acetylcholine receptor M5 protein (Bonner T.I. et al., Neuron 1:403-410 (1988)), and mouse a2A adrenoreceptor (Link R. et al., Mol. Pharmacol. 42:16-27 (1992)), respectively.

Example 2: The isolation of the human G protein-coupled receptor gene

[0046] The obtained rat sequence was subjected to EST search to reveal a fragment of the human homologue (gene bank NID: 946030 and NID: 901756). Human fetal brain cDNA was amplified by PCR using the specific primers IF01 (SEQ ID NO: 9/ CTTCCGCCGGGCCTTCACCAA) and IR02 (SEQ ID NO: 10/ ACAGACACGGCGGGGCTCAC) (probe 1). A human \(\tilde{L}\) EMBL3 SP6/T7 genomic library (Clontech) of a size of 1.2x10<sup>6</sup> pfu was screened using probe 1 according to standard procedure plaque hybridization procedures. Two positive clones were thus isolated. The obtained phage-clones were digested with SacI, and three bands of a clone were subcloned. These fragments, termed I1 (SEQ ID No: 11), I3 (SEQ ID No: 12) and I5 (SEQ ID No: 13), were sequenced and a hypothetical sequence was speculated by comparing with the rat homologue. I1 and I3 were subjected to PCR amplification using specific primers YS03 (SEQ ID NO: 14/ TGAACGCTTCGGGGGCGCTG) and YS05 (SEQ ID NO: 15/ GAGATGGCGAGGTTGAGCAGG), YS12 (SEQ ID NO: 16/ GGCTCCAAGCCATCGGCGTC) and YS14(SEQ ID NO: 17/ CTCACTTCCAGCAGTGCTCC) and the PCR products were termed probe 2 and probe 3, respectively. Human hypothalamus cDNA (1.3 x 10<sup>6</sup> phage) was plated at a density of 5.6 x 10<sup>4</sup> pfu/150mm plate. The obtained sub-pools were checked by PCR using the primers YS03 and YS05. One positive sub-clone was screened in the same method as the screening of the genomic library, using probe 2. One cDNA clone containing 5'UTR to TM5 was obtained and named cDNA clone 1.

[0047] Probe 4 was amplified by PCR from cDNA clone 1 using the primers YS07 (SEQ ID NO: 18/ GCCTCCG-CACCCAGAACAAC) and YS10 (SEQ ID NO: 19/ TGCGCCTCTGGATGTTCAG). Phase screening of the human hippocampus library (3x10<sup>6</sup> pfu) was done in the same method as the genomic library, using probe 3 and probe 4. A few clones were obtained and the longest one, termed cDNA clone 2, was sequenced. It has the region between TM2 and 3'UTR. cDNA clone 1 was digested by SacII, and the 3.3kb band, which contained vector and the 5'-end region, was treated by shrimp alkaline phosphatase. cDNA clone 2 was also digested by SacII, and the 1.7kb fragment was ligated into the 3.3kb fragment from cDNA clone 1. The clone into which this ligated fragment was inserted has been deposited at the National Institute of Bioscience and Human-Technology, Agency of Industrial Science and Technology.

Name of the depositary institution: National Institute of Bioscience and Human-Technology, Agency of Industrial Science and Technology, MITI.

Address of the depositary institution: 1-1-3 Higashi, Tsukuba, Ibaraki 305-8566, Japan.

Date of deposit: 12/17/1998. Accession Number: FERM BP-6609

[0048] Human "BG2" cDNA nucleotide sequence is shown in SEQ ID NO: 21, and the amino acid sequence of the protein encoded by the said cDNA in SEQ ID NO: 20.

[0049] Human "BG2" protein had 32%, 28%, and 27% homology to known G protein-coupled receptors; human  $\alpha$ -2C-1 adrenoreceptor (Regan J.W. et al., Proc. Natl. Acad. Sci. USA 85:6301-6305 (1988)), mouse  $\beta$ -1 adrenoreceptor (Jasper J.R. et al., Biochim. Biophys. Acta 1178:307-309 (1993)), and human muscarinic acetylcholine receptor M3 protein (Peralta E.G. et al., EMBO J. 6:3923-3929 (1987)), respectively.

#### Example 3: Northern blot analysis

[0050] Probe 4 was labeled with <sup>32</sup>Pγ-dCTP (Amersham, Prime It II) and used as cDNA probe for the detection of human "BG2". Human Multiple Tissue Northern (MTN) Blots Membrane with Express HybriTM was purchased from Clontech. After prehybridization of the membrane at 68°C for 30 min, it was hybridized with the probe at 68°C for 1 hr (final concentration of the probe was 1.5 x 10<sup>6</sup>cpm/ml.) The blot was rinsed with 2 x SSC containing 01% SDS at 42°C for 30 min, and the final wash was done at 50°C for 30 min in 0.1 x SSC containing 0.1% SDS. The blot was then exposed at -80°C for 2.5 days to Kodak autoradiographic film.

[0051] For the detection of the mouse "BG2," probe was prepared by PCR amplifying using the rat "BG2" cDNA as a template with sense primer MF2 (SEQ ID NO: 22/ TGCATCCCATTGTACGTNCC), and antisense primer MR1 (SEQ ID NO: 24/ TGCTCTGGGACACCATCTTC), purifying the amplified products by electrophoresis on an agarose gel, and labeling them as above described for human gene.

[0052] Blotting membrane used was the Rat MTN (Multiple Tissue Northern) blot (Clontech). Hybridization was performed at 42°C overnight in hybridization buffer (50% formamide, 4x SSPE, 1% SDS, 0.5% BLOTTO, and 100 µg/ml salmon sperm DNA). The membrane was washed at 65°C in 0.1x SSC containing 0.1% SDS, and then exposed to the Kodak autoradiography film at -80°C overnight.

[0053] The result showed that the human and rat "BG2" genes are strongly expressed particularly in the brain (Fig. 2).

### Example 4: In situ Hybridization

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[0054] Adult male Sprague-Dawley rats (Charles River Japan) aged 13-18 weeks were anesthetized with inhalation of ether, connected to a rotary pump and infused with chilled 4% paraformaldehyde in phosphate buffer (pH7.2) via a cannula inserted into the left ventricle. After perfusion, brain, pituitary and spinal cord were removed and dissected to sagittal or coronal sections. The tissue specimens were postfixed with the same fixative overnight at 4°C. The following process was carefully done to avoid RNA contamination. Tissue specimens were embedded in paraffin wax in a routine manner, then paraffin sections were cut into a thickness of 6 µm by rotary microtome (Model HM 355; MICROM Laborgerate GmbH). The sections were stored in moisture free condition at -20°C until proceeded to *in situ* hybridization.

[0055] For preparation of rat BG2 sense and antisense RNA probes, the cDNA fragment, amplified by PCR from MP-21 plasmid DNA using a sense primer MF2 (SEQ ID NO:22/ TGCATCCCATTGTACGTNCC) and antisense primer MR3 (SEQ ID NO:23/ ATCATTAGGAGCGTGTANGG), was cloned into pZErO-2 vector (Invitrogen). The RNA probes were labeled with digoxigenin using DIG RNA Labeling Kit (Boehringer Mannheim). Paraffin sections were de-paraffinized with xylene and transferred to distilled water after rinsing with graded ethyl alcohol. *In situ* Hybridization Reagents (ISHR, Code No. 316-01951; Nippon Gene) were used as reagents without digoxigenin-labeled RNA. The sections were incubated with two changes of phosphate buffer saline(PBS;ISHR 1) for 1 min and 10 min. The sections were treated with proteinase K (ISHR 6) for 10 min at 37°C. Acetylation was done with acetylation buffer (ISHR 3) containing acetic anhydride (ISHR 4) for 15 min, followed by quenching with PBS/glycine buffer (ISHR 2) for 20 min at room temperature, rinsed twice with 4 x SSC (ISHR 5) for 10 min and then rinsed with PBS buffer for 10 min. After pre-hybridization with 50% formamide/2 x SSC for 30 min at room temperature, hybridization was performed for 16 hr at 42 °C using digoxigenin-labeled RNA probe (1 μg/ml).

[0056] Post hybridization washing was performed twice 50% formamide/2 x SSC for 10 min at 42°C. Then the sections were treated with RNase A (ISHR10)/NET buffer (ISHR 9) for 30 min at 37°C after rinsing with NET buffer (ISHR 9) for 5 min at 37°C. After washing twice with 0.1 x SSC buffer (ISHR 11) for 20 min, the sections were transferred and labeled digoxigenin was detected using the Digoxigenin Detection Kit (Boehringer Manheim). Then the sections were rinsed with 100 mM Tris-Hcl, 150 mM NaCl containing buffer (Buffer 1) for 1 min at room temperature then incubated with blocking reagent (Buffer 2) for 30 min at room temperature. The sections were incubated with anti-digoxigenin alka-

line phosphatase-labeled antibody for 60 min at room temperature. After washing with Buffer 1 for 15 min and Buffer 3 for 2 min at room temperature, the solutions were incubated with NBT/X-phosphate solution diluted with Buffer 3 for 12-14 hr at room temperature. The sections were mounted with glycerol or Permount after washing with Buffer 4.

[0057] As a result, as shown in Figures 3 and 4, BG2 cDNA probe was strongly hybridized to the hippocampus and the spinal cord. Hybridization of a medium extent was also detected in hypothalamus and cerebellum.

#### Industrial Applicability

[0058] The present invention has provided novel G protein-coupled receptor proteins specifically expressed in the brain, and their genes. Use of the receptors makes it possible to screen their ligands and compounds that are candidates for medicines. These ligands and candidate compounds would be useful in the diagnosis and treatment of diseases arising from disorders of signal transduction pathway mediated by the G protein-coupled receptor of the invention.

### SEQUENCE LISTING

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. 15															•
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	Phe	e Let	ı Ser	r Val	Thr	Phe	Phe	Asr	Let	ı Ser	: Ile	Tyr	Leu	Ası	11e	Gln	
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	Trp Gly Cys Trp Pro Lys Gly His Gly Glu Ala Met Pro Leu His Ser	
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	teciggeege coageteteg geoggegeee tgeecegegt coeggageeg egtgageetg 30	) N
50	terrage condetered acondeter cancerated conducting the second	. •
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#### **Claims**

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- A guanosine triphosphate binding protein-coupled receptor protein comprising the amino acid sequence of SEQ ID NO: 1, or said amino acid sequence in which one or more amino acids are replaced, deleted, or added.
- A guanosine triphosphate binding protein-coupled receptor protein comprising the amino acid sequence of SEQ ID NO: 20, or said amino acid sequence in which one or more amino acids are replaced, deleted, or added.
- 3. A guanosine triphosphate binding protein-coupled receptor protein encoded by a DNA which hybridizes with a DNA comprising the nucleotide sequence of SEQ ID NO: 2.
- A guanosine triphosphate binding protein-coupled receptor protein encoded by a DNA which hybridizes with a DNA comprising the nucleotide sequence of SEQ ID NO: 21.
- 15 5. A partial peptide of the receptor protein as described in any one of claims 1 to 4.
  - A DNA encoding the receptor protein as described in any one of claims 1 to 4 or the partial peptide as described in claim 5.
- The DNA described in claim 6, wherein said DNA comprising the nucleotide sequence of SEQ ID NO: 2 or NO: 21.
  - 8. A vector containing the DNA as described in any one of claims 5 to 7.
  - 9. A transformant carrying the vector as described in claim 8.
  - 10. A method of producing the receptor protein as described in any one of claims 1 to 4 or the partial peptide as described in claim 5, the method comprising culturing the transformant as described in claim 9.
  - 11. A method of screening for a ligand of the receptor protein as described in any one of claims 1 to 4, the method comprising exposing a test compound to the receptor protein as described in any one of claims 1 to 4 or the partial peptide as described in claim 5, and selecting a compound that binds to said protein or partial peptide.
  - 12. A method of screening for a compound that inhibits the binding between the receptor protein as described in any one of claims 1 to 4 and its ligand, the method comprising,
    - (a) exposing a ligand to the receptor protein as described in any one of claims 1 to 4 or the partial peptide as described in claim 5 in the presence of a test compound, and detecting the binding activity between said protein or partial peptide and the ligand, and,
    - (b) comparing the binding activity detected in (a) with that in the absence of the test compound, and selecting a compound that reduces the binding activity between said protein or partial peptide and the ligand.
  - 13. A kit for screening a compound that inhibits the binding between the receptor protein as described in any one of claims 1 to 4 and its ligand, the kit comprising the receptor protein as described in any of claims 1 to 4 or the partial peptide as described in claim 5.
  - 14. An antibody which binds to the receptor protein as described in any one of claims 1 to 4.

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Figure 1

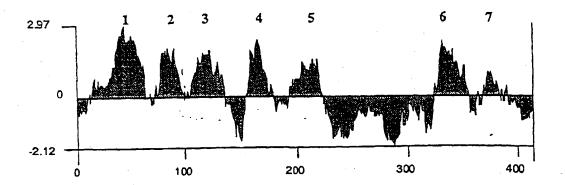
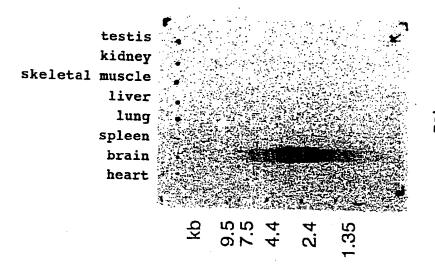
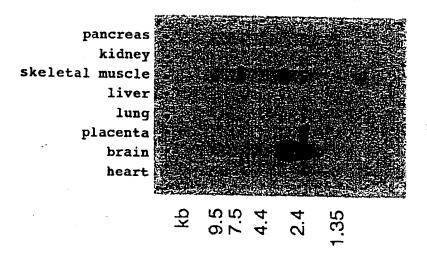


Figure 2





man

Figure 3

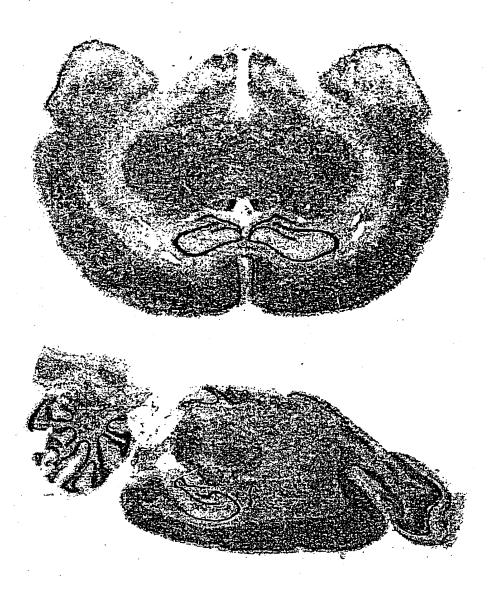
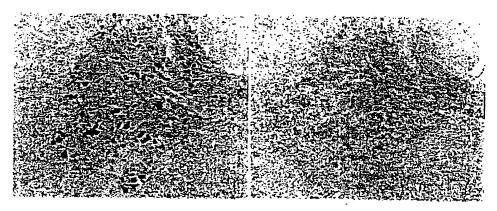


Figure 4



antisense

sense

### INTERNATIONAL SEARCH REPORT

International application No. PCT/JP98/05967

	CIFICATION OF SUBJECT MATTER C1 C12N15/12, C12N15/63, C12 C07K16/28, C12N1/21, C12N		14/705,	
According to	o International Patent Classification (IPC) or to both n			
B. FIELD	S SEARCHED			
	ocumentation searched (classification system followed C1 C12N15/12, C12N15/63, C12C07K16/28, C12N1/21, C12N	Q1/02, C12P21/08, C07K	14/705,	
Documentat	ion searched other than minimum documentation to th	e extent that such documents are included	d in the fields searched	
	ata base consulted during the international search (nat (DIALOG), BIOSIS (DIALOG), EMI			
C. DOCU	MENTS CONSIDERED TO BE RELEVANT			
Category	Citation of document, with indication, where ap		Relevant to claim No.	
Y	Bonner, T.I. et al., "Identif Muscarinic Acetylcholine Rec (1987) Vol. 237 p.527-532		1, 3, 5-14	
Y	Dewan, Z. et al., "Molecular characterization of a rat arufa2B-adrenergic receptor" Proc. Natl. Acad. Sci. USA (1990) Vol. 87, No. 8 p.3102-3106			
Y	JP,7-67654, A (Mitsubishi ) Laboratories, Inc.), 14 March, 1995 (14. 03. 95)		2, 4, 5-14	
Y	Ernest, G.P. et al., "Disting ligand-binding properties and expression of four human mus receptors" The ENBO J. (1987 p.3923-3929	d tissue-specific carinic acetylcholine	2, 4-14	
x Furthe	r documents are listed in the continuation of Box C.	See patent family annex.		
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International application No. PCT/JP98/05967

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